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(54) Method for determining state of disease progression.

(57) The present invention relates to a method for determining the state of disease progression thereby providing a means for the detection of said disease and in the risk evaluation for the development of said disease. In particular, the present invention contemplates a method for determining and monitoring the state of malignant or premalignant progression or risk of development of malignant or premalignant progression in mammalian tissue or cells. In particular, the present invention is directed to benign and malignant colon tumors and benignly and malignantly transformed colon cells. The present invention is predicated on the discovery that the abundance of specific first nucleic acids changes in response to the disease state. The present invention is also directed to recombinant second nucleic acids useful in the detection and quantification of the first nucleic acids by hybridization. The present invention also contemplates the use of polypeptides encoded by said second nucleic acids in the regression and prophylaxis of disease states.

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METHOD FOR DETERMINING STATE OF DISEASE PROGRESSION

The present invention relates to a method for determining the state of disease progression thereby providing a means for the detection of said disease and in the risk evaluation for the development of said disease. In particular, the present invention contemplates a method for determining and monitoring the state of malignant or premalignant progression or risk of development of malignant or premalignant progression in mammalian tissue and cells. Specifically, the present invention is directed to benign and malignant colon tumors and cells. The present invention is predicated on the discovery that the abundance of specific first nucleic acids changes in response to the disease state. The present invention is also directed to recombinant second nucleic acids useful in the detection and quantification of the first nucleic acids by hybridization. The present invention also contemplates the use of polypeptides encoded by said second nucleic acids in the regression and prophylaxis of disease states.

Of the 50,000 to 100,000 genes present in the genome in higher vertebrates, approximately 10,000 are expressed in any tissue at a particular development stage. The level of expression (mRNA concentration or abundance) of each of these varies widely from 1 copy per cell (rare) to 50-300 (middle abundant) to 1,000 (highly abundant) (Hastie and Bishop, 1976, Cell 9: 781). The large number of biochemical, enzymatic and antigenic differences between normal cells and their transformed counterparts indicates that changes in level of expression of many of these genes (aside from mutations, deletions, amplifications or other structural alterations) is involved in development of the fully malignant cell. Copending U.S. Patent Application No. 852,401 teaches methods to analyze the expression of a large representation of these genes in order to characterize tissue or cells as normal, benignly or malignantly transformed, or at risk for transformation or other phenotypes (e.g., responsiveness to various drug or biological therapeutic agents; potential for metastasis and likely site) which are of clinical importance.

The initial work utilized a dimethylhydrazine induced mouse colon tumor as a model (Augenlicht and Kobrin, 1982, Cancer Res. 42: 1088). A cDNA library of the expressed genes of this tumor was constructed and 400 random selections made. Standard methodology was used to determine relative levels of expression of each of these 400 sequences in a number of normal and neoplastic tissues. A semi-quantitative scale was used, and analyses were repeated a number of times. Several general conclusions were drawn. First, approximately 15% of the sequences changed in expression in the colon tumor as compared to the normal mouse colonic mucosa. Most of these (12%) were modest quantitative shifts. This extent of change is similar to that documented in a number of other systems of transformation, including rat hepatomas (Capetanaki and Alonso, 1980, Nuc. Acids Res. 8: 3193; Jacobs and Birnie, 1980, Nuc. Acids Res. 8: 3087), human lymphoid neoplasia (Hanania, et al., 1981, Proc. Natl. Acad. Sci., USA, 78: 6504), and most important, even the relatively well understood transformation of primary chick embryo fibroblasts by the Rous sarcoma virus (RSV) (Groudine and Weintraub, 1980, Proc. Natl. Acad. Sci., USA, 77: 5351). Hence, even when the etiology of transformation is well understood (e.g., the introduction of the src gene by RSV and the expression of its product, pp80 sarc), the cell rapidly exhibits a large number of changes in gene expression which may include alterations in as many as 1000 sequences. Among those sequences whose normal expression was relatively restricted to the colon, there were many decreases (nineteen) in expression in colon tumors as well as modest increases (twenty three). Fewer changes (nine) were seen in the tumors among those sequences expressed in other normal tissues, but the alterations were of much larger magnitude (Augenlicht and Kobrin, supra).

In moving to the human, several significant changes were made. First, the number of sequences from a reference cDNA library made from the HT-29 human colon carcinoma cell line was increased to 4,000 (Augenlicht, et al., 1987, Cancer Res., 47: 6017). This provided an 80% probability that every abundant and middle abundant sequence in this colon carcinoma cell line was represented in the data set. Second, methods were developed to accomplish the analysis of expression of each of these 4,000 sequences in very small human biopsies which yield 50-100 ng of poly A+ RNA. Utilizing a computerized scanning and image processing system, (Augenlicht, et al., supra; U.S. Patent Appl. No. 852,401) relative level of expression of each of the 4000 sequences was quantitated in each biopsy. The number of sequences screened was reduced from 4000 in a series of experiments. First, all 4000 clones were evaluated in two biopsies of normal mucosa from individuals at low genetic risk for colon cancer; in two biopsies of benign adenoma from patients with the autosomal dominant disease familial polyposis; and in two biopsies of two different colon carcinomas. Sequences which were expressed at near background levels (low abundance) in all six of these biopsies, or which were modestly above background and showed no evidence of alteration in level of expression among the tissues, were eliminated. This left 378 clones for additional screening with other biopsies (Augenlicht, et al., supra).

Several facts emerged from the large data base generated. First, the overall number of sequences which changed in expression between human colon carcinomas and normal human colonic mucosa was approximately 7%, which is the same order of magnitude as the extent of change seen in other systems cited above. Furthermore, the change was progressive, in that fewer alterations were seen in benign tumors (adenomatous polyps) than in malignant carcinomas when both were compared to normal tissue - the flat mucosa from patients at low genetic risk for colon cancer. Finally, the flat mucosa from patients with familial polyposis who are at very high risk for development of colon cancer showed much greater changes in gene expression when compared to low-risk normal mucosa than either the benign polyps that arise in this disease or the carcinomas. Hence, the high-risk tissue, having undergone many constitutive alterations in association with the inherited gene defect, may be primed for progression along any of many pathways to malignancy.

This same methodology and reference library was then used to analyze changes in gene expression in HT-29 and SW-480 colon carcinoma cell lines induced to differentiate in vivo with sodium butyrate. Again, many alterations in gene expression were found, but a comparison of the in vivo and in vitro data bases allowed selection of eight sequences whose relative levels of expression characterized colonic cells as either differentiated or fully transformed. Further, the quantitative extent of change in these sequences in in vivo and in vitro was similar, with a linear correlation coefficient which was significant for a comparison of the in vivo and in vitro data at the p 0.01 level. The in vivo data establish that the in vitro results are not tissue culture artifacts but do in fact bear a relationship to the human disease. Conversely, the in vitro data could be confirmed by standard Northern blot analysis, thus validating the scanning and image processing methodology, and also reduce the complexity of cell types and human tissue variability for further analysis of these sequences.

In accordance with the present invention, it has been discovered that expression of one of the aforementioned sequences can be used to determine the state of benign and malignant colon tumors as compared to the normal colonic mucosa, thus fulfilling an important need to monitor such tumors.

The present invention contemplates a method for determining the state of malignant or premalignant progression or risk for development of malignant or premalignant progression in mammalian tissues and cells which comprises identifying a statistically significant differential in detected abundance of an RNA isolated from the tissue to be studied relative to a predetermined standard, ie., abundance value obtained from tissue isolated from a particular population.

The present invention is also directed to a recombinant nucleic acid molecule useful in the detection of the aforementioned RNA.

Another aspect of the present invention relates to a polypeptide encoded by said recombinant nucleic acid molecule.

Still another aspect of the present invention contemplates a method for regression and inhibition of growth of malignant and benign colonic tumors by the administration of an effective amount of the aforementioned polypeptide with a pharmaceutically acceptable carrier.

In the accompanying drawings,

Figure 1(A) is a bar diagram depicting expression of 50F1 in biopsies taken from normal mucosa (M), adenomas (A) and colon carcinomas (C).

Figure 1(B) is a graphical representation of expression of 50F1 in tissue culture (in vitro) following treatment of HT29 colon carcinoma cells with sodium butyrate.

Figure 2 is a photographic representation of a Northern blot analysis of RNA from HT29 cells from 12 hours to 96 hours exposed to sodium butyrate. "C" represents a control.

Figure 3 is a photographic representation of a Southern blot analysis of genomic DNA from different tissues hybridized with 50F1.

Figure 4 is a bar diagram depicting copy number of 50F1 in different tissue samples.

Figure 5 is a bar diagram depicting copy number of 50F1 in different tissue samples at different stages of development.

Figure 6 is a bar diagram depicting copy number of 50F1 in different tissue samples.

Figure 7 is a diagrammatic representation of the cDNA clone 50F1 showing nucleotide sequence and partial restriction map.

The present invention contemplates a method for determining the state of malignant or premalignant progression or risk for development of malignant or premalignant progression in mammalian tissues and cells which comprises identifying a statistically significant differential in detected abundance of an RNA isolated from the tissue to be studied relative to a standard. The selection of said standard being determined by the analysis being conducted. For example, if cancer states are to be detected in a random

population, the standard is a predetermined abundance obtained from a normal population (i.e., free from detectable cancer and genetically at low risk). If the analysis is to monitor the effectiveness of an anticancer treatment, said standard is a predetermined abundance obtained from an anormal (cancerous or benignly transformed) population. In one preferred embodiment, the mammalian tissue is human colonic mucosa but the skilled artisan will recognize the applicability of the method disclosed herein to a wide variety of tissue such as breast, pancreatic, stomach, lung, oral, cerebral, intestinal and the like.

More particularly, the present invention contemplates a method for determining the state of malignant or premalignant progression or risk for development of malignant progression in human colonic mucosa or colon cells which comprises determining the relative abundance of an RNA, said RNA being represented in a first nucleic acid isolated from the tissue to be studied and immobilized onto a solid support, by contacting to said first nucleic acid, a second nucleic acid in probe form, said second nucleic acid characterized by possessing a copy number variably represented in different normal human tissues, such that the second nucleic acid, or portion thereof, will hybridize to said RNA, the hybridization quantified by exposing said second nucleic acid contacted with said RNA to a detecting means.

The present invention is predicated on the surprising discovery that a group of nucleic acid sequences, characterized herein as possessing copy numbers variably represented in different normal mammalian tissue compared to other nucleic acid sequences which are relatively constant in their representation in the same tissue. In accordance with the present invention, a simple dot-blot method is described enabling the rapid determination of the variable or constant copy number of a particular sequence.

In a specific embodiment, a nucleic acid sequence possessing a copy number which is variably represented is designated herein 50F1, isolated from human colonic mucosa and is further defined by its nucleotide sequence given in Fig. 7. In accordance with the present invention, expression of 50F1 progressively decreases in benign and malignant colon tumors in humans as compared to the normal colonic mucosa (Fig. 1A) and increases back to the level characteristic of normal mucosa when HT29 colon carcinoma cells *in vitro* are induced to differentiate by exposure to 5mM sodium butyrate (Fig. 1B). Northern blot analysis of RNA from HT29 cells confirms this increase in expression at 24 to 96 hours following treatment with sodium butyrate (Fig. 2).

Southern blot analysis demonstrates that the copy number of this sequence is elevated in the genomic DNA isolated from the colon carcinoma cell lines HT29 and SW480 and in genomic DNA from two colon tumors relative to the level in the DNA from two normal human placentas (Fig. 3).

Accordingly, one of the many benefits of the present invention is the discovery of a nucleic acid sequence useful in determining the state of malignant or premalignant progression or risk of development of malignant or premalignant progression. As previously indicated, many such nucleic acid sequences are contemplated which may be useful in the practice of this invention. The present invention is disclosed using one particular sequence (50F1) which, up to the present time, has been determined to be most useful in practicing this invention. This is done, however, with the understanding that the present invention encompasses all such sequences.

For example, in accordance with the subject invention, a series of experiments was carried out in which genomic DNA was isolated from a number of normal human tissues and hybridized to one of the aforementioned nucleic acid sequences, designated herein 50F1. Level of hybridization, and hence, relative copy number of the sequence, was determined by careful attention to quantitation and standardization of the hybridization results. This was done as follows. A measured amount of each DNA sample (approximately 1 micrograms) was applied to nitrocellulose in duplicate at 2 positions using a standard 96 position (8 x 12 format) dot-blot apparatus (Schleicher and Schuell, Inc.). The duplicate dots were not adjacent on the dot-blot. They were placed at sites distal to each other to avoid potential systematic error due to position effects during application, hybridization, washing, or film exposure. Similarly, several positions throughout the dot-blot were left blank for background correction. The source of the DNA samples varied (below), but every blot contained duplicate positions of DNA from placenta J which therefore provided a standard between experiments. Finally, every dot-blot was made in replicate so that one copy could be hybridized to the sequence of interest (e.g., 50F1) and a replicate to a standard (e.g., human β -globin). This was used to correct for potential differences in the amount of DNA loaded per dot for each sample.

A 32 P labeled probe was made using the insert of either 50F1 or human β -globin using the well known nick-translation procedure and each hybridized to a replicate of a dot-blot. Following washing, the blots were exposed to X-ray film to obtain a visual image of the relative hybridization of each probe to each DNA sample.

For quantitation, the following procedure was used. The dot-blot was cut into sections, each section being the location of one dot (i.e., one DNA sample). Each section was then placed into liquid scintillation

fluid and the extent of hybridization of the probe to the DNA determined by recording the counts per minute for each section in a scintillation counter (Packard Tricarb).

Calculation was then done as follows. The mean hybridization for each DNA sample, done at least in duplicate on a blot was calculated. Similarly, the mean background was calculated from the blank spots. The mean background for a blot was then subtracted from the mean hybridization for each DNA sample on that blot. For each DNA sample, a corrected value was determined by calculating the ratio: mean hybridization 50F1/mean hybridization β -globin, the latter data being determined in the same way from the replica blot which had been hybridized to a human β -globin probe and having been standardized by reference to placenta J on that blot. Finally, standardization between experiments was established by taking this calculated 50F1 value for each DNA sample and expressing it as a ratio to the value calculated similarly for the DNA sample placenta J, which was included on every blot.

In summary, these calculations correct for differences in determination of amount of DNA applied to the dot-blot, and differences between experiments (e.g., probe specific activity, conditions of hybridization).

Experiments done and calculated in the manner described have yielded the following results. For purposes of description, differences in hybridization level between genomic DNA of different samples or tissues are referred to as differences in copy number of the sequence (i.e., 50F1) in the genome.

Fig. 4 illustrates that relative to the mean of 6 different placentas, the copy number of 50F1 is elevated three to nine fold in 3 colon carcinoma cell lines (HT29, SW480 and SKCO1). A three fold elevation was also seen in genomic DNA from colon tumors and a similar elevation was seen in the normal human colonic mucosa. In Fig. 5, the range for the 6 placentas is shown as 2 horizontal lines (approximately 0.8-1.6). Fig. 5 illustrates that the copy number differs over a wide range among different tissues and at different developmental stages (gestational age of the fetus). No difference is detected among the placentas, peripheral blood leukocytes (PBL), bone marrow, and spleen at any developmental stage. Stomach shows a slight elevation at 32 weeks, somewhat higher in the adult. The three fold elevation in colon is essentially established by 22 weeks. No increase is seen in diaphragm at 22 weeks, but this rapidly increases to seven fold at 32 weeks and 12 fold in the adult. Liver, kidney, and adrenal similarly show progressive increases in 50F1 copy number with developmental stage.

The data in Fig. 6 extends this analysis to other adult tissues including: thymus, prostate, lymph node, pancreas, thyroid, bladder, lung, heart, spinal cord, psoas muscle, and perirenal fat pad. A range of copy numbers of 50F1 is seen among these tissues.

Similar experiments utilizing either a v-Kirsten-ras probe or C-myc probe instead of 50F1 demonstrated that these sequences do not vary in copy number among human fetal and adult tissues.

The data of Fig. 1 illustrated the utility of evaluation of level of expression of sequence 50F1 for distinguishing between normally differentiated and benign and malignant transformed colonic cells *in vivo* and *in vitro*. The data shown demonstrate that the copy number of 50F1 is elevated in normal colonic tissue, and varies among tissues and developmental stages, indicating that 50F1 is important in the processes of differentiation and transformation and clinically useful in characterizing the differentiation or transformation status of human colonic, and other, epithelial cells. Furthermore, the subject invention contemplates a method for determining the effectiveness of a material in regressing or inhibiting growth of colonic cancer comprising identifying a statistically significant differential in detected abundance of an RNA isolated from mammalian tissue or cells, said tissue having been subjected to contact with said material for a time sufficient to effect regression or inhibition of growth of colonic cancer, said differential being relative to a standard. Said standard being selected from a predetermined abundance obtained from normal or abnormal populations.

The present invention also relates to polypeptides, or parts thereof, encoded by nucleic acid sequences, or parts thereof, wherein said nucleic acid sequences are characterized as possessing a copy number variably represented in different mammalian tissue. For example, the present invention is directed to the polypeptide or its derivatives (e.g., precursor) encoded by 50F1 and to the larger polypeptide encoded by the sequences adjacent to 50F1 in the genome.

The desired polypeptides are synthesized *in vivo* by first determining a nucleic acid sequence (RNA or DNA) encoding the amino acid sequencing comprising said polypeptide, inserting said nucleic acid sequence into an expression vector, transforming the resulting recombinant molecule into a suitable host and then culturing or growing the transformed host under conditions requisite for the synthesis of the polypeptide. The recombinant molecule defined herein should comprise a nucleic acid sequence encoding a desired polypeptide inserted downstream of a promoter, a eukaryotic or prokaryotic replicon and a selectable marker such as resistance to an antibiotic. The recombinant molecule may also require a signal sequence to facilitate transport of the synthesized polypeptide to the extracellular environment. Alternatively, the polypeptide may be retrieved by first lysing the host cell by a variety of techniques such as

sonication, pressure disintegration or toluene treatment. Hosts contemplated in accordance with the present invention can be selected from the group comprising prokaryotes (e.g., *Escherichia coli*, *Bacillus* sp., *Pseudomonas* sp.) and eukaryotes (e.g., mammalian cells, yeast and fungal cultures, insect cells and plant cultures). The artisan will also recognize that a given amino acid sequence can undergo deletions, substitutions and additions of nucleotides or triplet nucleotides (codons). Such variations are all considered within the scope of the present invention. Techniques useful in practicing this aspect of the invention can be found in Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, pages 1-500, 1982.

Alternatively, once the amino acid sequence is known, the polypeptide can be synthesized chemically (i.e., *in vitro*). In one example, a solid phase methodology of synthesis can be used starting with a resin to which the amino acid residue located at the amino end of the molecule is linked and to which subsequent amino acids, are selectively added. A resin commonly used in the art is benzhydrylamine (BHA resin) which is derived from a cross-linked polystyrene bead resin manufactured by copolymerization of styrene and divinylbenzene. Resin of this type is known and its preparation is further demonstrated by Pletta, et al., Chem. Commun. 650; 1970 and Orłowski, et al., J. Org. Chem. 41: 3701, 1976. In this synthesis, the amino acids are added one at a time to the insoluble resin until the total polypeptide sequence has been built up on the resin. The functional groups of the amino acids are protected by a blocking group. Blocking groups are well known in the art. For example, the alpha amino group of the amino acids can be protected by a tertiary butyloxycarbonyl group. The hydroxyl functions of serine and threonine can be protected by a benzyl or benzyl derivative group such as 4-methoxybenzyl, 4-methylbenzyl, 3,4-dimethylbenzyl, 4-chlorobenzyl, 2,6-dichlorobenzyl, 4-nitrobenzyl, benzhydryl or an equivalent thereof. The first amino acid residue (amino end) is coupled to the BHA resin by standard techniques such as under amide forming conditions. The blocked amino acid residue BHA produced above can then be deprotected by washing in a solvent with gentle agitation.

Selected amino acids are then singularly added. At the end of the synthesis, the polypeptide is cleaved from the resin by, for example, treatment with hydrogen fluoride and then the polypeptide is purified using standard known techniques such as HPLC. The artisan will recognize variation in the aforementioned method as well as alternative synthesis techniques. All these are considered within the scope of the present invention.

The polypeptides contemplated herein are useful in the regression and palliation of some cancers and tumors, and in particular, colonic tumors and cancers. This is especially evident with the polypeptide encoded by 50F1 and related sequences since it is evident that less of this polypeptide will be present in hosts with advanced malignant progression. Accordingly, the subject invention contemplates a method for inducing regression or inhibiting growth of colonic cancer by administering a pharmaceutical composition containing an effective amount of said polypeptide. Additionally, a method for inducing regression or inhibiting growth of colonic cancer in a mammal is contemplated in which a nucleic acid molecule encoding the polypeptides contemplated herein is introduced into an affected (i.e., cancerous or transformed) cell in such a manner that said nucleic acid molecule is expressed in the cytoplasm of said cell or following integration into the genome of said cell.

In this case, the nucleic acid molecule is carried to said affected cell and transferred into said cell by a second nucleic acid molecule (e.g., various viruses). The first nucleic acid molecule is manipulated such that it contains the appropriate signals for expression. That is, in accordance with the present invention, a method of inducing regression or inhibiting growth of colonic cancer in a mammal is contemplated comprising administering a first nucleic acid molecule encoding the aforementioned polypeptide, said nucleic acid being contained in a pharmacologically acceptable second nucleic acid carrier molecule such that said first nucleic acid enters a target cell and is either maintained in extrachromasomally or integrates into the genome of said target cell in such a manner that said first nucleic acid is expressed so as to produce an effective amount of said polypeptide.

The active ingredients of the pharmaceutical compositions comprising said polypeptide, are contemplated to exhibit excellent and effective therapeutic activity, for example, in the treatment of some colonic cancers. Thus, the active ingredients of the therapeutic compositions and the novel compounds of the present invention which act as agonists or antagonists for endogenous peptides or non-peptide drugs when administered in amounts up to about 1000 mg per kilogram of body weight per day. This dosage regimen may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation. A decided practical advantage is that the active compound may be administered in a convenient manner such as by the oral, intravenous (where water soluble), intramuscular or subcutaneous routes.

The active compounds may be orally administered, for example, with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard or soft shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet. For oral therapeutic administration, the active compound may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 1% of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 5 to about 80% of the weight of the unit. The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained. Preferred compositions or preparations according to the present invention are prepared so that an oral dosage unit form contains up to about 1000 mg of active compound.

The tablets, troches, pills, capsules and the like may also contain the following: A binder such as gum tragacanth, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such as sucrose, lactose or saccharin may be added or a flavoring agent such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. A syrup or elixir may contain the active compound, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavoring such as cherry or orange flavor. Of course, any material used in preparing a dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may be incorporated into sustained-release preparations and formulations.

The active compounds may also be administered parenterally or intraperitoneally. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredient into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from previously sterile-filtered solution thereof.

As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect in association with the

required pharmaceutical carrier. The specification for the novel dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active material and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active material for the treatment of disease in living subjects having a diseased condition in which bodily health is impaired as herein disclosed in detail.

The principal active ingredient is compounded for convenient and effective administration in effective amounts with a suitable pharmaceutically acceptable carrier in dosage unit form as hereinbefore disclosed. A unit dosage form can, for example, contain the principal active compound in amounts ranging from 1 to about 1000 mg. Expressed in proportions, the active compound is generally present in from about 1 to about 1000 mg/ml of carrier. In the case of compositions containing supplementary active ingredients, the dosages are determined by reference to the usual dose and manner of administration of the said ingredients.

The present invention also relates to antibodies to the aforementioned polypeptides. Such antibodies are contemplated to be useful in developing detection assays (immunoassays) for said polypeptides, especially during examination of biopsy tissue and in the purification of these polypeptides. The antibodies may be monoclonal or polyclonal. Additionally, it is within the scope of this invention to include any second antibodies (monoclonal or polyclonal) directed to the first antibodies discussed above. The present invention further contemplates use of these second antibodies in detection assays and, for example, in monitoring the effect of an administered pharmaceutical preparation. Furthermore, it is within the scope of the present invention to include antibodies to the glycosylated regions of said polypeptides, and to any molecules complexed with said polypeptides. Accordingly, in accordance with this invention, an antibody to said polypeptides encompasses antibodies to the polypeptide, or part thereof, and to any associated molecules (e.g., glycosylated regions, lipid regions, carrier molecules, and the like).

The polypeptides, or parts thereof, considered herein are purified then utilized in antibody production. Both polyclonal and monoclonal antibodies are obtainable by immunization with the polypeptides, and either type is utilizable for immunoassays. The methods of obtaining both types of sera are well known in the art. Polyclonal sera are less preferred but are relatively easily prepared by injection of a suitable laboratory animal with an effective amount of the purified polypeptide, or part thereof, collecting serum from the animal, and isolating specific sera by any of the known immunoabsorbent techniques. Although antibodies produced by this method are utilizable in virtually any type of immunoassay, they are generally less favored because of the potential heterogeneity of the product.

The use of monoclonal antibodies in the present immunoassay is particularly preferred because of the ability to produce them in large quantities and the homogeneity of the product. The preparation of hybridoma cell lines for monoclonal antibody production derived by fusing an immortal cell line and lymphocytes sensitized against the immunogenic preparation can be done by techniques which are well known to those who are skilled in the art. (See, for example, Douillard, J. Y. and Hoffman, T., "Basic Facts About Hybridomas", in Compendium of Immunology, Vol. II, L. Schwartz (Ed.) (1981); Kohler, G. and Milstein, C., Nature 256: 495-497 (1975); European Journal of Immunology, Vol. 6, pp. 511-519 (1976); Koprowski, et al., U.S. Patent 4,172,124, Koprowski et al., U.S. Patent 4,196,265 and Wands, U.S. Patent 4,271,145, the teachings of which are herein incorporated by reference.

Unlike preparation of polyclonal sera, the choice of animal is dependent on the availability of appropriate immortal lines capable of fusing with lymphocytes thereof. Mouse and rat have been the animals of choice in hybridoma technology and are preferably used. Humans can also be utilized as sources for sensitized lymphocytes if appropriate immortalized human (or nonhuman) cell lines are available. For the purpose of the present invention, the animal of choice may be injected with from about 1 mg to about 20 mg of the purified polypeptide, or part thereof. Usually the injecting material is emulsified in Freund's complete adjuvant. Boosting injections may also be required. The detection of antibody production can be carried out by testing the antisera with appropriately labeled antigen. Lymphocytes can be obtained by removing the spleen or lymph nodes of sensitized animals in a sterile fashion and carrying out fusion. Alternatively, lymphocytes can be stimulated or immunized in vitro, as described, for example, in C. Reading J. Immunol Meth., 53: 261-291 1982.

A number of cell lines suitable for fusion have been developed, and the choice of any particular line for hybridization protocols is directed by any one of a number of criteria such as speed, uniformity of growth characteristics, deficiency of its metabolism for a component of the growth medium, and potential for good fusion frequency.

Intraspecies hybrids, particularly between like strains, work better than interspecies fusions. Several cell lines are available, including mutants selected for the loss of ability to secrete myeloma immunoglobulin. Included among these are the following mouse myeloma lines: MPC₁₁-X45-6TG, P3-NS1-1-Ag4-1, P3-X63-

Ag8, or mutants thereof such as X83-Ag8.653, SP2-0-Ag14 (all BALB/C derived), Y3-Ag1.2.3 (rat), and U266 (human).

Cell fusion can be induced either by virus, such as Epstein-Barr or Sendai virus, or polyethylene glycol. Polyethylene glycol (PEG) is the most efficacious agent for the fusion of mammalian somatic cells. PEG itself may be toxic for cells, and various concentrations should be tested for effects on viability before attempting fusion. The molecular weight range of PEG may be varied from 1,000 to 8,000. It gives best results when diluted to from about 20% to about 70% (w/w) in saline or serum-free medium. Exposure to PEG at 37°C for about 30 seconds is preferred in the present case, utilizing murine cells. Extremes of temperature (i.e., above 45°C) are avoided, and preincubation of each component of the fusion system at 37°C prior to fusion gives optimum results. The ratio between lymphocytes and malignant cells is optimized to avoid cell fusion among spleen cells and a range of from about 1:1 to about 1:10 gives good results.

The successfully fused cells can be separated from the myeloma line by any technique known by the art. The most common and preferred method is to choose a malignant line which is Hypoxanthine Guanine Phosphoribosyl Transferase (HGPRT) deficient, which will not grow in an aminopterin-containing medium used to allow only growth of hybrids and which is generally composed of hypoxanthine 1×10^{-4} M, aminopterin 1×10^{-5} M, and thymidine 3×10^{-5} M, commonly known as the HAT medium. The fusion mixture can be grown in the HAT-containing culture medium immediately after the fusion 24 hours later. The feeding schedules usually entail maintenance in HAT medium for two weeks and then feeding with either regular culture medium or hypoxanthine, thymidine-containing medium.

The growing colonies are then tested for the presence of antibodies that recognize the antigenic preparation. Detection of hybridoma antibodies can be performed using an assay where the antigen is bound to a solid support and allowed to react to hybridoma supernatants containing putative antibodies. The presence of antibodies may be detected by "sandwich" techniques using a variety of indicators. Most of the common methods are sufficiently sensitive for use in the range of antibody concentrations secreted during hybrid growth.

Cloning of hybrids can be carried out after 21-23 days of cell growth in selected medium. Cloning can be performed by cell limiting dilution in fluid phase or by directly selecting single cells growing in semi-solid agarose. For limiting dilution, cell suspensions are diluted serially to yield a statistical probability of having only one cell per well. For the agarose technique, hybrids are seeded in a semisolid upper layer, over a lower layer containing feeder cells. The colonies from the upper layer may be picked up and eventually transferred to wells.

Antibody-secreting hybrids can be grown in various tissue culture flasks, yielding supernatants with variable concentrations of antibodies. In order to obtain higher concentrations, hybrids may be transferred into animals to obtain inflammatory ascites. Antibody-containing ascites can be harvested 8-12 days after intraperitoneal injection. The ascites contain a higher concentration of antibodies but include both monoclonals and immunoglobulins from the inflammatory ascites. Antibody purification may then be achieved by, for example, affinity chromatography.

The presence of the polypeptides contemplated herein, or antibodies specific for same, in a patient's serum or biopsy tissue can be detected utilizing antibodies prepared as above, either monoclonal or polyclonal, i.e., virtually any type of immunoassay. A wide range of immunoassay techniques are available as can be seen by reference to U.S. Patent Nos. 4,016,043, 4,424,279 and 4,018,653. This, of course, includes both single-site and two-site, or "sandwich", assays of the non-competitive types, as well as in traditional competitive binding assays. Sandwich assays are among the most useful and commonly used assays and are favored for use in the present invention. A number of variations of the sandwich assay technique exist, and all are intended to be encompassed by the present invention. Briefly, in a typical forward assay, an unlabeled antibody is immobilized in a solid substrate and the sample to be tested brought into contact with the bound molecule. After a suitable period of incubation, for a period of time sufficient to allow formation of an antibody-antigen binary complex, a second antibody, labeled with a reporter molecule capable of producing a detectable signal is then added and incubated, allowing time sufficient for the formation of a ternary complex of antibody-labeled antibody. Any unreacted material is washed away, and the presence of the antigen is determined by observation of a signal produced by the reporter molecule. The results may either be qualitative, by simple observation of the visible signal, or may be quantitated by comparing with a control sample containing known amounts of hapten. Variations on the forward assay include a simultaneous assay, in which both sample and labeled antibody are added simultaneously to the bound antibody, or a reverse assay in which the labeled antibody and sample to be tested are first combined, incubated and then added to the unlabeled surface bound antibody. These techniques are well known to those skilled in the art, and then possibly of minor variations will be readily

apparent.

In the typical forward sandwich assay, a first antibody having specificity for the polypeptide, or part thereof, contemplated in this invention is either covalently or passively bound to a solid surface. The solid surface is typically glass or a polymer, the most commonly used polymers being cellulose, polyacrylamide, nylon, polystyrene, polyvinyl chloride or polypropylene. The solid supports may be in the form of tubes, beads, discs or microplates, or any other surface suitable for conducting an immunoassay. The binding processes are well-known in the art and generally consist of cross-linking covalently binding or physically adsorbing the molecule to the insoluble carrier. Following binding, the polymer-antibody complex is washed in preparation for the test sample. An aliquot of the sample to be tested is then added to the solid phase complex and incubated at 25° C for a period of time sufficient to allow binding of any subunit present in the antibody. The incubation period will vary but will generally be in the range of about 2-40 minutes. Following the incubation period, the antibody subunit solid phase is washed and dried and incubated with a second antibody specific for a portion of the hapten. The second antibody is linked to a reporter molecule which is used to indicate the binding of the second antibody to the hapten. By "reporter molecule," as used in the present specification, is meant a molecule which, by its chemical nature, provides an analytically identifiable signal which allows the detection of antigen-bound antibody. Detection may be either qualitative or quantitative. The most commonly used reporter molecules in this type of assay are either enzymes, fluorophores or radionuclide containing molecules (i.e., radioisotopes). In the case of an enzyme immunoassay, an enzyme is conjugated to the second antibody generally by means of glutaraldehyde or periodate. As will be readily recognized, however, a wide variety of different conjugation techniques exist, which are readily available to the skilled artisan. Commonly used enzymes include horseradish peroxidase, glucose oxidase, B-galactosidase and alkaline phosphatase, among other. The substrates to be used with the specific enzymes are generally chosen for the production, upon hydrolysis by the corresponding enzyme, of a detectable color change. For example, p-nitrophenyl phosphate is suitable for use with alkaline phosphatase conjugates; for peroxidase conjugates, 1,2-phenylenediamine, 5-aminosalicylic acid, or toluidine are commonly used. It is also possible to employ fluorogenic substrates, which yield a fluorescent product rather than the chromogenic substrates noted above. In all cases, the enzyme-labeled antibody is added to the first antibody hapten complex, allowed to bind, and then the excess reagent is washed away. A solution containing the appropriate substrate is then added to the ternary complex of antibody-antigen-antibody. The substrate will react with the enzyme linked to the second antibody, usually visual signal, which may be further quantitated, usually spectrophotometrically, to give an indication of the amount of hapten which was present in the sample.

Alternately, fluorescent compounds, such as fluorescein and rhodamine, may be chemically coupled to antibodies without altering their binding capacity. When activated by illumination with light of a particular wavelength, the fluorochrome-labeled antibody absorbs the light energy, inducing a state of excitability in the molecule, followed by emission of the light at a characteristic color visually detectable with a light microscope. As in the EIA, the fluorescent labeled antibody is allowed to bind to the first antibody-hapten complex. After washing off the unbound reagent, the remaining ternary complex is then exposed to the light of the appropriate wavelength, the fluorescence observed indicates the presence of the hapten of interest. Immunofluorescence and EIA techniques are both very well established in the art and are particularly preferred for the present method. However, other reporter molecules, such as radioisotope, chemiluminescent or bioluminescent molecules, may also be employed. It will be readily apparent to the skilled technician how to vary the procedure to suit the required purpose. It will also be apparent that the foregoing can be used to detect directly or indirectly (i.e., via antibodies) the polypeptides of this invention.

Plasmid 50F1 is maintained in a culture of *Escherichia coli* which has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland on April 14, 1988 under the accession number 67674. The deposit was made in accordance with the provision to the Budapest Treaty. Said culture will remain permanently available for a term of at least five years after the most recent request for the furnishing of a sample and, in any case, 30 years after the date of the deposit. Should the culture become nonviable or be inadvertently destroyed, it will be replaced with a viable culture of the same taxonomic description.

The following examples further illustrate the present invention.

EXAMPLE 1

Preparation of RNA

Biopsy samples of human colonic mucosa, colonic adenomas and carcinomas are taken using a flexible
 5 colonoscope and placed into liquid nitrogen within one minute of removal. The samples are stored in liquid
 N₂ until their use. RNA is prepared from the biopsy material by rapid disruption of the tissue in a buffer
 containing guanidinium isothiocyanate and pelleting the RNA through a cushion of CsCl as described by
 Maniatis, *et al.*, supra. PolyA + RNA is isolated by chromatography on oligo dT cellulose by the method of
 Aviv and Leder, supra. The PolyA + RNA is used to prepare ³²P labeled cDNA probes as described by
 10 Augenlicht and Kobrin, supra.

EXAMPLE 2

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Scanning the Films and Data Digitization

20 Exposed x-ray film following hybridization of the library in a format is scanned with an Eikonix scanner,
 model 785, supported by a VAX 11/780 computer (Digital Equipment Corporation). The data are subse-
 quently analyzed and processed using an IP 8500 image processing system (Gould) supported by the VAX
 11/780 and an AP 120B x-ray processor (floating point system).

The Eikonix scanner records signal from the S-ray films in areas called pixels. The signal consists of
 25 the % transmitted light and is collected using a 45mm wide angle lens. It is then converted to optical
 density for each pixel. The pixel size is set at 100 x 100 microns, and the film is therefore divided into a
 grid 2048 x 2048 pixels for a total of 4.19×10^6 individual measurements for each film (11 films = 1
 complete replica of the library hybridized to one probe). Each of these values represents an 8 bit optical
 density value obtained from a 12 bit transmission value.

30 The resultant image is first processed with a 5x5 median filter to suppress grain and other high
 frequency noise. This procedure replaces each pixel with the median value of the pixels in the 5x5
 neighborhood surrounding it. Background is then subtracted in two steps. An intermediate image is
 produced from the median filtered image by choosing the minimum value of a 161 pixel horizontal line
 centered at the element being generated. The actual length of the line is not critical as long as it is wider
 35 than the region reserved for an individual clone. A second intermediate image is produced from the first in
 the same manner except that a maximum value is used in place of the minimum value. This image is then
 subtracted on a pixel by pixel basis to produce the processed image from which clone measurements are
 made.

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EXAMPLE 3

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Analysis of 50F1

The cDNA clone 50F1 was analyzed with respect to its nucleotide sequence and restriction cleavage
 pattern (restriction map).

50 A portion of the nucleotide sequence of the molecule cloned in the plasmid termed 50F1 is shown in
 Fig. 7 along with a partial restriction map of this portion of the plasmid. The sequence is shown as the
 reverse complement to present the open reading from position 45 to 227. Hence, the homopolymeric G
 sequence at the beginning of the sequence is located at the right end of the restriction map. This
 homopolymeric region was put on the end of the molecule by in vitro manipulation for cloning.

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EXAMPLE 4

Hybridization Studies With 50F1

50F1, a 500bp cDNA clone, is useful as a probe for screening human genomic DNA for related
 5 sequences from adult and fetal tissue, including colon, placentas and HT29 cells and primary colon tumors.
 Using the methods contained herein, the 50F1 sequence is detected in high molecular weight DNA and,
 following digestion, in a single fragment whose size depends on the restriction enzyme used.

Rigorous quantitation of DNA dot blots indicates that, in contrast to human B-globin, c-myc and c-k-
 ras2 which remain constant, the copy number of 50F1 is elevated 6 fold in HT29 cells exposed to Na
 10 butyrate and an average of 3 fold in colon tumors relative to the DNA from several different placentas. A
 similar elevation is present in normal colon as early as 22 weeks of gestation. Further, the copy number
 varies amongst and is characteristic of different tissues and developmental stage. Thus, 50F1 is a sequence
 whose relative level of expression reflects the differentiation/ transformation state of colon cells in vivo and
 in vitro and whose genomic copy number changes during development in the normal colon and other
 15 normal human tissues, suggesting an important role in normal differentiation.

Claims

- 20 1. A method for determining and monitoring the state of malignant or premalignant progression or risk
 for development of malignant or premalignant progression in mammalian tissues which comprises identify-
 ing statistically significant differential in detected abundance of an RNA isolated from the tissue to be
 studied relative to a standard.
- 25 2. The method according to Claim 1 wherein said RNA is represented in a first nucleic acid isolated
 from said tissue and immobilized onto a solid support.
3. The method according to Claim 1 wherein a second nucleic acid, or part thereof, will hybridize to
 said RNA, or part thereof, said second nucleic acid characterized by possessing a copy number variably
 represented in different normal mammalian tissues.
- 30 4. The method according to Claim 3 wherein the hybridization is quantified by exposing said second
 nucleic acid contacted with said RNA to a detecting means.
5. A method for determining the state of malignant or premalignant progression or risk for development
 of malignant or premalignant progression in mammalian tissue or cells as defined in Claim 1 comprising
 determining the relative abundance of an RNA, said RNA being represented in a first nucleic acid isolated
 from the tissue to be studied by contacting to said first nucleic acid, a second nucleic acid in probe form.
 35 said second nucleic acid characterized by possessing a copy number variably represented in different
 normal mammalian tissues, such that the second nucleic acid, or part thereof, will hybridize to said RNA,
 the hybridization quantified by exposing said second nucleic acid contacted with said RNA to a detecting
 means.
6. The method according to Claim 5 wherein said mammalian tissue is human tissue.
- 40 7. The method according to Claim 6 wherein said human tissue is derived from human colonic mucosa.
8. The method according to Claim 5 wherein said first nucleic acid comprises total cellular RNA.
9. The method according to Claim 5 wherein the second nucleic acid probe is labeled with a material
 which generates a signal, said material being selected from the group consisting of a radioactive isotope, an
 enzyme, a fluorescent molecule, a chemiluminescent molecule and a bioluminescent molecule.
- 45 10. The method according to Claim 5 wherein the detecting means is selected from the group
 consisting of x-ray film, an enzyme substrate and a device for detecting luminescence.
11. The method according to Claim 5 wherein said nucleic acid is complementary DNA (cDNA) to RNA
 derived from malignant tissue.
12. The method according to Claim 11 wherein said cDNA is maintained in a culture of Escherichia coli,
 50 accorded the ATCC accession number 67674, and designated 50F1.
13. The method according to Claim 11 or 12 wherein said cDNA is characterized by the following
 nucleotide sequence:

10 20 30 40 50
 AA GGG GGG GGG GGG GGG GGG GGG GGG CAC ACA TCA ATC ACC TGA GGT CAC CAT AGT CTA
 5 60 70 80 90 100 110
 ATA GAA AAC AAC CGA AAC CAA ATG AGA TTC AAG CAC TGC TGC ATT ACA ATT GTT ACT
 120 130 140 150 160 170
 10 GGG TCT CTA TTT ACC CTC CCT ACA AGC CTC AGA GTA CTT CGA GTC TCC CTT CAC ATT
 180 190 200 210 220 230
 GAT CGA CGG CAT CTA CGG CTC AAC ATT TTT TGT AGC CAA GGC TTC AGG ATT GAG TA

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14. A cDNA characterized by the following nucleotide sequence:

10 20 30 40 50
 20 AA GGG GGG GGG GGG GGG GGG GGG GGG CAC ACA TCA ATC ACC TGA GGT CAC CAT AGT CTA
 60 70 80 90 100 110
 ATA GAA AAC AAC CGA AAC CAA ATG AGA TTC AAG CAC TGC TGC ATT ACA ATT GTT ACT
 25 120 130 140 150 160 170
 GGG TCT CTA TTT ACC CTC CCT ACA AGC CTC AGA GTA CTT CGA GTC TCC CTT CAC ATT
 180 190 200 210 220 230
 30 GAT CGA CGG CAT CTA CGG CTC AAC ATT TTT TGT AGC CAA GGC TTC AGG ATT GAG TA

15. The cDNA according to Claim 14 wherein said cDNA is contained in a vector.
16. The cDNA according to Claim 15 wherein the vector is selected from the group consisting of a plasmid, a cosmid and a viral vector.
17. The cDNA according to Claim 16 wherein said vector is capable of replicating in a eukaryotic host or a prokaryotic host.
18. The cDNA according to Claim 16 wherein the vector is pBR322.
19. The cDNA according to Claim 17 wherein the host is selected from the group consisting of bacteria, mammalian cells, yeast cells, fungal cells and insect cells.
20. A host transformed with the cDNA according to Claim 18 or 19.
21. The host according to Claim 20 wherein said host has the identifying characteristics of *Escherichia coli* and accorded the ATCC accession number 67674.
22. A polypeptide encoded by the cDNA according to Claim 14.
23. A pharmaceutical composition useful in inducing regression or inhibiting growth of colonic cancer in a mammal comprising an effective amount of polypeptide according to Claim 22 and a pharmacologically acceptable carrier.
24. A pharmaceutical composition useful in inducing regression or inhibiting growth of colonic cancer in a mammal comprising an effective amount of first nucleic acid according to Claim 5 and a pharmacologically acceptable second nucleic acid carrier molecule.
25. An antibody to the polypeptide, or a part thereof, according to Claim 22.
26. The antibody according to Claim 25 wherein said antibody is a monoclonal or a polyclonal antibody.
27. A method for detecting the polypeptide according to Claim 22 in human colonic mucosa comprising the steps of contacting the tissue or tissue extract to be tested with an antibody to said polypeptide, or part thereof, for a time sufficient for a polypeptide-antibody complex to form and subjecting said polypeptide-antibody complex to a detecting means.

28. The method according to Claim 27 wherein said antibody is labelled with a material providing a detectable signal, said material selected from the group consisting of a radioactive isotope, an enzyme, a fluorescent molecule, a chemiluminescent molecule or a bioluminescent molecule.

29. The method according to Claim 27 or 28 wherein the polypeptide-antibody complex is detected by
5 a second antibody, said second antibody being directed to the first antibody, wherein said second antibody being labelled with a detectable signal.

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FIG.IB

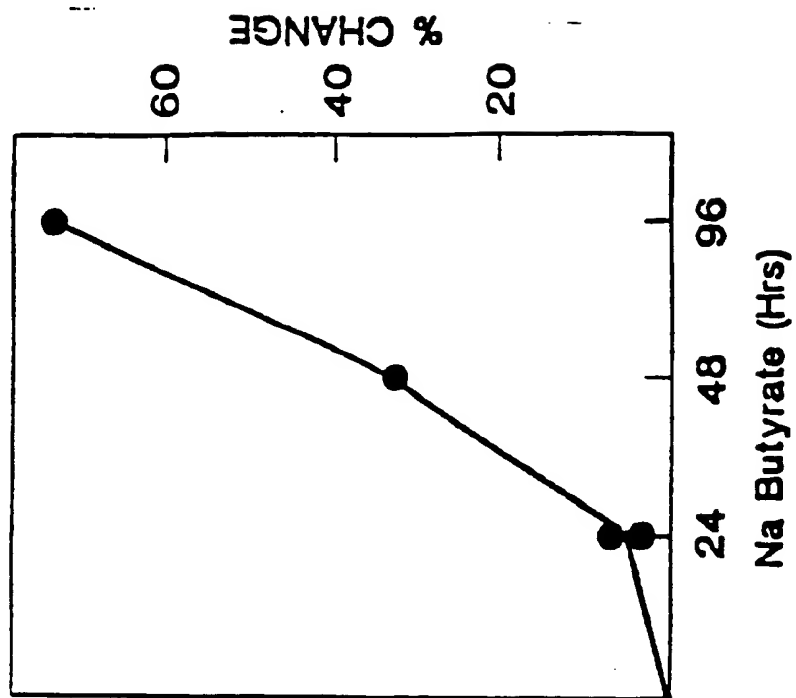
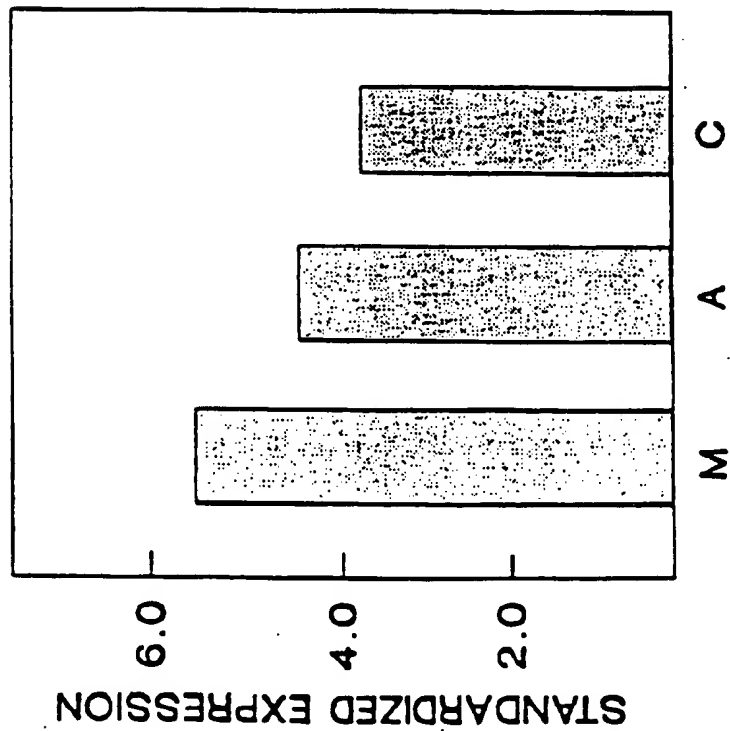


FIG.IA



C 12 24 48 96


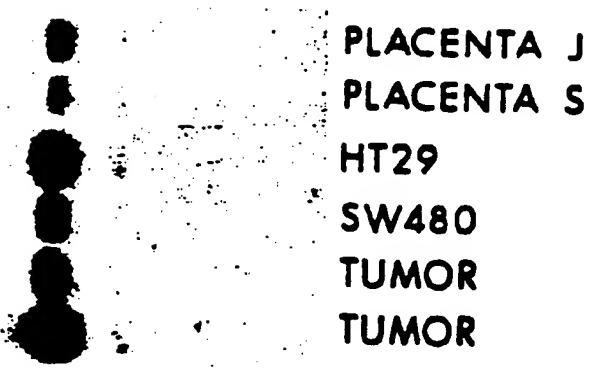


FIG.2



FIG.3



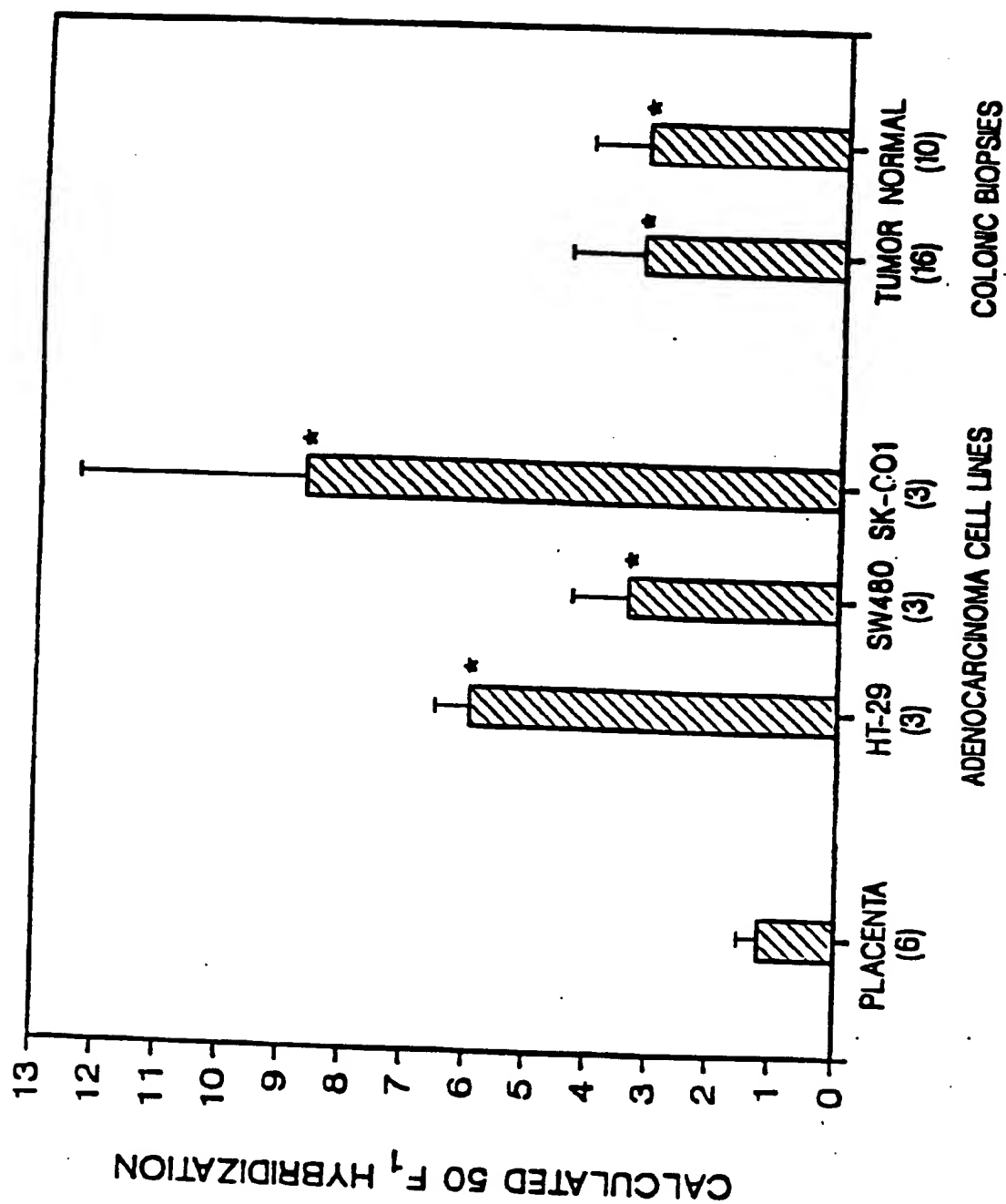


FIG.5

